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Effect of Foliar Application of Free Amino Acids on Salt Tolerance and Yield Quality of Manzanillo Olive Tree

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Abstract: This study aimed to investigate the impact of different types of free amino acids (proline, glutamic acid, and GABA), applied either individually or in combination, on the vegetative growth and yield of nineyear-old Manzanillo olive trees over two consecutive seasons (2021 and 2022) in a private orchard located along the Cairo-Alexandria desert road, Egypt (approximately 64 km from Cairo). The findings revealed that applying a mixture of free amino acids at a rate of 50 ppm positively influenced vegetative growth, flowering, fruit physical properties, and yield, with no significant differences compared to foliar application of 50 ppm glutamic acid alone. Additionally, the combined amino acid treatment enhanced the olive trees' resistance to salt stress by increasing total amino acid content in the leaves while reducing carbohydrate content.

Key words: Olive, amino, proline, glutamic, gaba, productivity.

1. Introduction

The olive tree (Olea europaea L.) is among the most economically significant fruit crops globally, valued for its oil and table olives. However, its cultivation faces numerous challenges, particularly abiotic stresses, with salinity being a major concern, especially in arid and semi-arid regions. Salt stress adversely affects plant physiology by causing osmotic imbalance, oxidative damage, and ion toxicity, ultimately resulting in diminished growth, yield, and quality. (Lodolini et al., 2019; Al-Maaitah et al., 2017 and Nogueira et al., 2022). Amino acids, as crucial components of plant metabolism, have garnered attention for their potential to ameliorate salt stress effects in olive trees.

The purpose of this review is to present a comprehensive understanding of the uses of amino acids to improve salt tolerance in olive cultivation and to identify future research directions for optimizing their application in practical agricultural settings. The use of foliar-applied amino acids to mitigate these adverse effects,

enhancing the resilience and productivity of olive trees in saline environments. This review

summarizes recent research findings on the role of amino acids in improving salt tolerance and yield quality in olive cultivation (**Zrig** *et al.*, **2023** and **Di Vaio** *et al.*, **2024**).

Recent research by Zhang and Zhou (2020) and Kumar and Singh (2020) has highlighted the roles of amino acids in maintaining ion homeostasis and synthesizing stress-responsive metabolites, respectively. Studies by Ben Ahmed *et al.* (2020) and Zrig *et al.* (2023) have provided valuable insights into how amino acids influence hormonal signaling and metabolic adjustment in olive trees subjected to saline environments. Amino acids serve as precursors for the synthesis of stress-responsive metabolites and osmoprotectants in olive trees. They participate in the biosynthesis of compatible solutes, polyamines, and secondary metabolites that have a role in the mechanisms of stress tolerance (El-Tayeb, 2021; Liu *et al.*, 2022 and El-Esawi and Ali 2022).

Amino acids such as proline and glycine betaine have been shown to accumulate in plants under salinity stress. These compounds act as osmoprotectants, helping cells maintain turgor pressure and preventing ion imbalance by stabilizing cellular structures. Recent studies highlight that proline application leads to a significant reduction in oxidative damage by modulating osmotic adjustment and ion transport (Ghars *et al.*, 2022).

Amino acids contribute to osmotic adjustment by accumulating in plant cells, thereby maintaining turgor pressure and water uptake under saline conditions. Proline, glycine betaine, and arginine are among the most prominent amino acids implicated in osmotic regulation in olive trees subjected to salt stress (Iqbal *et al.*, 2021; Farooq *et al.*, 2020; Orlandi *et al.*, 2021 and Hassan and Ali 2022).

Salinity stress triggers the production of reactive oxygen species (ROS), causing oxidative damage in plants. Amino acids such as cysteine and methionine serve as precursors for key antioxidants like glutathione. Applying these amino acids as foliar sprays enhances the activity of antioxidant enzymes, including superoxide dismutase (SOD) and peroxidase (POD), thereby mitigating oxidative stress and enhancing the plant's tolerance to salinity. (Hasanuzzaman *et al.*, 2020; Ahmad and Sharma 2021; El-Esawi *et al.*, 2020; Zrig *et al.*, 2023 and Mosleh & Soltani, 2023).

One key aspect of salinity stress is the disruption of ion balance within plant tissues. Exogenous application of amino acids enhances the selective absorption of essential ions like potassium (K+) while limiting the accumulation of toxic ions like sodium (Na+). Excessive sodium uptake under salt stress disrupts ion homeostasis in olive plants. Amino acids play a role in ion compartmentalization and sequestration, thereby minimizing the toxic effects of sodium and maintaining ion balance within cells (Ali *et al.*, 2021 and Hamed *et al.*, 2021). Recent research in 2023 revealed that foliar application of amino acid blends markedly enhances nutrient uptake efficiency, promoting improved growth in plants under saline conditions. (Zrig *et al.*, 2023 and Di Vaio *et al.*, 2024). Cysteine, glutamine, and asparagine are implicated in ion homeostasis mechanisms in olive trees exposed to saline environments (Hu *et al.*, 2015; Zhang and Zhou 2020 and Wang *et al.*, 2022).

Amino acids like tryptophan are precursors for plant hormones such as auxins. Studies show that foliar application of tryptophan increases auxin levels, promoting root development and improving water and nutrient uptake. This hormonal modulation helps maintain vegetative growth and yield even in saline environments (Mosleh and Soltani, 2023). Amino acids influence hormonal signaling pathways involved in plant stress responses. Amino acid blends regulate the biosynthesis, perception, and signaling of stress-related hormones, including abscisic acid (ABA), cytokinins, and jasmonates, in olive trees. This modulation helps coordinate adaptive responses to salt stress (Zhang and Jiang 2021; Fariduddin *et al.*, 2021; Khan and Müller 2022).

Recent studies from 2022 and 2023 emphasize the beneficial effects of amino acid foliar sprays on olive fruit yield and quality under salinity stress. These findings indicate that such applications enhance oleic acid content and improve oil stability, both of which are essential quality attributes for olive oil (**Ghars** *et al.*, **2022**). Salinity stress often leads to poor fruit set and reduced yields. Amino

acid treatments, particularly those including proline, arginine, and glutamine, have been linked to increased fruit retention, improved chlorophyll content, and higher overall yields (**Zrig** *et al.*, **2023**). Amino acid treatments contribute to better nutrient translocation and cell enlargement, resulting in improved fruit size and weight even under stress conditions (**Di Vaio** *et al.*, **2024**).

This study aimed to investigate the impact of individual free amino acids and a mixture of three amino acids on salt tolerance and yield performance of Manzanillo olive trees.

2. Materials and Methods

A field study was conducted over two consecutive growing seasons (2021 and 2022) on 25 nine-year-old Manzanillo olive trees grown in sandy soil under a drip irrigation system. The trees, spaced 4 x 6 meters apart, were located in a private orchard 64 kilometers from Cairo, at coordinates 30°268'215" N latitude and 30°806'534" E longitude. The study aimed to investigate the role of amino acids in enhancing olive tree growth and yield. All selected trees were disease-free, uniform in size and shape, and irrigated with groundwater. Standard cultural practices and fertilization were applied. Soil physical and chemical characteristics, along with water chemical properties, were analyzed by the Soil, Water, and Environmental Research Institute of the Agricultural Research Centre, using the methods described by **Wilde** *et al.* (1985). The results of these analyses are summarized in Tables 1 and 2.

			Ph	ysical	anal	yses (%)				
	Coarse sand		Fine sand			Silt	Cl	lay	Texture class	
	38.4		43.3	43.3		13.0	5.3		Sandy	
	Chemical analyses (Anions and Cations) mg/L									
pH	ECds /m	Ca++	Mg^{++}	Na	+	\mathbf{K}^{+}	CO	HCO ₃ -	Cl.	SO 4
7.99	0.56	1.8	0.87	2.5	5	1.25		1.51	4.51	0.43
	Available nutrients (mq/ Kg Soil)									
	N	Р		K						
12	7.3	15.8	99	9.8						

Table (1). Physical and chemical analysis of the experimental soil

Table (2). Chemical characteristics of the tested water sample collected from the experimental area

	Soluble cations(me/L)				Soluble anions (me/ L)					
рН	E.C. ds/M (1:5)	Ca ⁺⁺	Mg^{++}	Na^+	\mathbf{K}^{+}	CO	HCO ₃ -	Cl-	SO4	SAR
7.12	6.10	22.69	16.54	16.54	0.17	-	1.52	18.75	47.91	6.25

The study was designed in a complete randomized block design with three replicates, two trees were used as a separator between treatments. The selected trees received the following foliar spray treatments during swelling buds stage and before the beginning of flowering:

1- Control (sprayed with water only).

- 2- Glutamic amino acid at 50 ppm.
- 3- Proline amino acid at 50 ppm.
- 4- GABA amino acid at 50 ppm.
- 5- A mixture of previous amino acids at 50 ppm.

The Following Parameters were estimated:

2.1. Vegetative Growth Measurements

Measurements such as shoots length (cm), No. of leaves/shoot and Leaf area (cm²) were made in August of each season on sixteen healthy shoots (4 in each tree direction) which selected randomly and labeled. Leaf area (cm²) were measured by using the planimeter.

2.2. Flowering Behavior

The following behaviors were estimated and recorded: No. of inflorescence/shoot, No. of flower/ inflorescence, sample of inflorescences were randomly taken at bloom stage from the middle portion of shoot from each replicate and No. of flower were counted. Sex ratio % it was calculated (Sex ratio % = perfect flower/ total No. of flower * 100).

2.3. Fruit physical characteristics

Physical characteristics of olive fruits such as Fruit shape index (fruit length/ fruit diameter), seed shape index (seed length/ seed diameter) and Pulp % (pulp weight/ fruit weight * 100) were determined at harvesting time.

2.4. Yield

The number of fruits per shoot, fruits per meter, and the average yield per tree (kg/tree) were recorded at harvest time (October) for each treatment during both growing seasons. Fruit weight was measured using an electric balance, with samples consisting of 10 fruits per replicate. The average fruit weight was then calculated. The oil content (%) of dried fruit samples was determined using petroleum ether (60–80%) as a solvent over 16 hours with a Soxhlet apparatus, following the procedure outlined by A.O.A.C. (1998).

2.5. Chemical Analysis

- Total amino acids

The total amino acids in olive leaves was determined according to Yemm and Cocking (1995).

- Total carbohydrates

Total sugars (carbohydrates) were determined according to the method of Dubois et al. (1956).

2.6. Data Analysis

The obtained data were statistically analyzed (Senedecor and Cochran, 1990). Duncan s multiple range tests effect was used to compare treatment means (Duncan, 1955).

3. Results and discussion

3.1. Vegetative growth

Data in Table (3) showed a significant difference among different treatments during two studied seasons. the highest significant shoot length (cm) were recorded by all amino acid treatments. While the least shoot length (17.29 - 17.33 cm) came from the control treatments in both seasons respectively.

Regarding the number of leaves per shoot, data presented in Table (3) indicate that amino acid treatments did not produce any significant effect during the first season. However, in the second season, the highest number of leaves per shoot was observed in the control treatment, recording 53.42 leaves.

Also, data indicate that leaf area of Manzanillo olive cultivar were ranged from $(3.29 - 4.60 \text{ cm}^2)$ in the 1st season and from $(3.55 - 4.94 \text{ cm}^2)$ in the second one. The highest leaf area was recorded by the foliar spray with a mixture of amino acids in the 1st and 2nd seasons, respectively.

The positive effect of amino acids on vegetative growth of olive tree may explained by its function on protein synthesis and the formation of enzymes important for the metabolism process (Aberg, 1961 and Mervat *et al.*, 2015). Additionally, Davies (1982) and Feng *et al.* (2020) stated that amino acids help regulate cellular pH due to their buffering capacity and metabolic roles. This regulation is critical for maintaining an environment conducive to enzymatic reactions and cellular metabolism. Moreover, in case the plant faces a deficiency of carbohydrates, Amino acids give plant cells an instant source of nitrogen and can also serve as a source of carbon and energy (Goss, 1973: Thon *et al.*, 1981, Calvo *et al.*, 2014 and Zayed *et al.*, 2023).

Tucotmont	Shoot ler	ngth (cm)	No. of	leaves	Leaf area (cm ²)		
Treatment	1 st	2 nd	1 st	2 nd	1 st	2 nd	
Control	17.29 ^в	17.33 ^C	38.42 ^A	53.42 ^A	3.90 ^в	4.00 ^C	
Glutamic	20.48 ^A	20.25^{ABC}	37.08 ^A	43.92 ^в	4.04 ^B	4.24 ^B	
Proline	18.64 ^{AB}	18.12 ^{BC}	43.67 ^A	45.50 ^B	3.29 ^C	3.55 ^D	
GABA	19.58 ^{AB}	22.04 ^A	38.92 ^A	40.33 ^B	4.14 ^B	4.16 ^{BC}	
Mix	19.97 ^{AB}	20.75^{AB}	37.58 ^A	45.00 ^B	4.60 ^A	4.94 ^A	

Table (3).	Effect	of	foliar	spray	with	amino	acids	on	vegetative	growth	of	olive	trees	cv.
	Manzai	nillo	o on tw	o studi	ed sea	sons (2	021 an	d 20	22)					

Values sharing the same letter are not significantly different at the 5% significance level.

The obtained results are in harmony with those stated by **Psarras** *et al.* (2024) on olive and **El-Badawy** (2019) on apricot. **Nafea and Al-Janabi** (2021) illustrated that olive shoot length increased significantly by the using of amino acid. In addition, **El-Bolok and Kasem** (2023) observed an enhancement on the vegetative growth of olive trees c.v Agazzi when it sprayed with chitosan at 1% + amino acids at 2%.

3.2. Flowering Behavior

The data in Table (4) reveal that olive flowering behavior was gradually increased by using amino acids. As such, the foliar spray with Glutamic amino acid gave the highest significant No. of inflorescences/ shoot, No. of flower/inflorescence and sex ratio % in two studied seasons without significant differences with those treated by the mixture of tested amino acids in most cases.

Amino acids have the capacity to regulate plant stress responses and this reflected in improving floral behavior. They have the ability to regulate the levels of important phytohormones like salicylic acid and abscisic acid (ABA), which are involved in plant development, stress resistance, and flowering. In addition, **Psarras** *et al.* (2024) illustrated that, No. of inflorescences of olive 'Koroneiki cv.' were increased significantly when it treated by amino acid spray.

Table (4). Effect of foliar spray with amino acids on flowering b	behavior of olive trees cv.
Manzanillo on two studied seasons (2021 and 2022)	

Tracting and	No of infl	orescence	No. of	flower	Sex ratio		
Treatment	1 st	2 nd	1 st	2 nd	1 st	2 nd	
Control	6.25 ^в	11.83 ^B	11.83 ^B	15.00 ^A	47.09 ^{AB}	66.50 ^C	
Glutamic	9.75 ^A	14.33 ^A	13.67 ^A	15.33 ^A	58.11 ^A	86.39 ^{AB}	
Proline	8.67 ^{AB}	7.58 ^C	12.58 ^{AB}	14.33 ^A	37.45 ^в	74.62 ^C	
GABA	7.58 ^{AB}	9.25 ^C	12.92 ^{AB}	14.17 ^A	51.45 ^{AB}	83.71 ^B	
Mix	10.00 ^A	12.00 в	13.67 ^A	15.00 ^A	43.52 ^B	93.98 ^A	

Values sharing the same letter are not significantly different at the 5% significance level.

3.3. Fruiting and fruit character

No. of fruit/shoot

The results in Table (5) indicate that the application of amino acids significantly increased the No. of fruits per shoot in treated plants compared to the untreated control. In the 1st season, the highest No. of fruits per shoot was recorded with the spraying of Glutamic amino acid, resulting in 5.92 fruits per shoot. In the 2nd season, a mixture of amino acids, including Glutamic acid, Proline, and Gapa, resulted in the highest No. of fruits per shoot at 7.67. These findings suggest that amino acid treatments, particularly Glutamic and the amino acid mix, positively influenced fruit production across both seasons. In the same order of amino acid treatments, regardless of the variance between them, the lowest mean No. of fruits per shoot was observed under Proline treatment, which recorded 3.08 fruits per shoot in the 1st season. Meanwhile, in the 2nd season, the control (untreated) treatment showed the lowest value, with 4 fruits per shoot. This further emphasizes that while amino acid treatments generally increased fruit production, Proline was the least effective in the 1st season, and untreated plants had the lowest yield in the 2nd season.

The effectiveness of the Glutamic acid in enhancing fruit production aligns with research indicating glutamic acid plays a significant role in plant growth and development. Glutamic acid has been shown to improve various plant physiological processes, which can enhance fruit characteristics (**Eisaa** *et al.*, **2023**). Similarly, the amino acid mix's superior performance in the 2nd season supports findings that combining amino acids can synergistically benefit plant growth and fruit yield (**Liu** *et al.*, **2020**).

In contrast, Proline treatment yielded the lowest No. of fruits per shoot in the 1st season (3.08 fruits), which suggests that Proline alone may be less effective in promoting fruit production compared to other amino acids or combinations. The control treatment also showed the lowest fruit production in the 2nd season (4 fruits per shoot), highlighting the overall benefits of amino acid treatments. This observation is consistent with studies that emphasize the advantages of amino acid supplementation over no treatment (**Ekşi and Sönmez, 2022**).

Number of fruits/meter

The results presented in (Table 5) illustrated that the application of various amino acids significantly affected the mean No. of fruits per meter. Spraying with amino acids resulted in a substantial increase in the No. of fruits per meter compared to the control. There was significant variance among the treatments with Glutamic, Proline, Gapa, and the amino acid mix across both seasons. The highest values were recorded with the Glutamic treatment, achieving 31.80 fruits per meter in the 1st season and 36.55 in the 2nd season. Additionally, the amino acid mix (Glutamic, Proline, and Gapa) recorded the highest value in the 2nd season at 38.75 fruits per meter. These findings highlight the positive impact of Glutamic and the amino acid mix on fruit yield. The lowest mean values of fruits per meter were observed with Proline (16.61) in the 1st season and in the control treatment (23.5) during the 2nd season. This suggests that while amino acid treatments generally enhanced fruit yield per meter, Proline was the least effective in the 1st season, and untreated plants had the lowest fruit yield in the 2nd season.

This result supports the role of Glutamic in promoting fruit production. Glutamic acid improves plant tolerance by enhancing water retention, reducing oxidative stress, and increasing antioxidant enzyme activity. Also, it boosts proline levels for osmotic balance and ABA accumulation, activating drought-resistance pathways that strengthen cellular defenses and reduce damage under stress, (**Xu**, *et al.*, **2020**) these actions can lead to increased fruit yield. Furthermore, the amino acid mix, which includes Glutamic acid, Proline, and Gapa, achieved the highest fruit yield in the 2nd season (38.75 fruits per meter). This finding is consistent with studies suggesting that combinations of amino acids can have synergistic effects on plant growth and fruit production (Liu *et al.*, **2020**).

The significant variance among treatments indicates that not all amino acids had the same impact on fruit yield. Proline treatment yielded the lowest No. of fruits per meter in the 1st season (16.61 fruits), and the control treatment recorded the lowest fruit yield in the 2nd season (23.5 fruits). Proline, while beneficial in some contexts, was less effective compared to glutamic and the amino acid mix, which might be due to its limited role in enhancing overall fruit production under the conditions tested. The control's lower fruit yield further underscores the advantages of amino acid supplementation over no treatment.

Fruit shape index

Table (5) shows that the application of all amino acid treatments led to an increase in the fruit shape index compared to the untreated plants, which had the lowest values (1.11 and 1.16 as the mean for both seasons). The greatest fruit shape index was observed under the glutamic amino acid treatment, with mean values of 1.30 in the 1^{st} season and 1.36 in the 2^{nd} season. This indicates that glutamic amino acid treatment had the most significant effect on improving the fruit shape index across both seasons.

The significant improvement in fruit shape index with glutamic amino acid treatment aligns with existing literature that highlights the role of amino acids in fruit development. glutamic acid is known to influence various plant physiological processes, including growth and stress responses. Its application can enhance cellular metabolism and improve overall fruit quality, including shape and uniformity (**Brosnan and Brosnan, 2013; Almutairi** *et al., 2022*). The superior performance of glutamic in improving the fruit shape index could be attributed to its role in supporting root growth, calcium transport, and hormonal balance.

Seed shape index

The results presented in Table (5) indicate a significant effect of amino acid treatments on the seed shape index in both seasons. The foliar spray with glutamic amino acid produced the highest seed shape index values, with 1.83 in the 1st season and 1.92 in the 2nd season. The differences among the other treatments were insignificant, further highlighting the superior influence of glutamic amino acid on seed shape index compared to the other amino acids.

Pulp %

Table (5) illustrated the pulp percentage (%), showing that the mix of amino acids was the most effective treatment, resulting in 87.18% and 86.26% pulp in the 1st and 2nd seasons, respectively. The lowest pulp percentage was recorded in the control, Glutamic, and Gapa amino acid treatments during the 1st season. While there were no significant differences between most treatments, the amino acid mix demonstrated a notably superior effect on pulp percentage across both seasons.

This finding aligns with previous research suggesting that amino acid treatments can enhance various fruit characteristics, including pulp quality. For instance, studies have shown that mixtures of amino acids can synergistically enhance plant growth and fruit development by optimizing nutrient uptake and physiological processes (Ekşi and Sönmez, 2022). This observation suggests that while individual amino acids can improve certain fruit characteristics, their effects on pulp percentage might not be as pronounced as those achieved with a combination of amino acids. Glutamic acid and Gapa have been reported to affect plant metabolism and stress responses (Zhang and Jiang, 2021), but their impact on pulp percentage might be limited compared to a well-balanced amino acid mixture. It is possible that the effects of individual amino acids or specific combinations may vary depending on the plant species, growth conditions, and fruiting stage.

Treatment	No fruit/	. of shoot	No fruit/i	. of meter		shape lex	Seed s ind	-	Pul	p %
	1 st	2 nd	1 st	2 nd	1 st	2 nd	1 st	2 nd	1 st	2 nd
Control	3.83 ^B	4.00 ^D	22.73 ^A B	23.50 ^B	1.11 ^C	1.16 ^C	1.78 ^A в	1.76 c	84.28 c	84.20 ^B
Glutamic	5.92 ^A	7.25 ^A B	31.80 ^A	36.55 ^A	1.30 ^A	1.31 ^A B	1.83 ^A	1.82 в	83.67 c	83.68 B
Proline	3.08 ^B	4.75 ^C	16.61 ^B	27.49 ^B	1.26 ^A B	1.36 ^A	1.76 ^B	1.85 B	85.53 в	86.26 A
Gapa	4.67 ^A B	6.08 ^B C	23.37 ^A B	31.54 ^A B	1.25 ^A B	1.34 ^A B	1.75 ^B	1.84 в	84.02 c	84.57 ^B
Mix	4.08 ^B	7.67 ^A	19.89 ^в	38.75 ^A	1.22 ^в	1.27 ^в	1.77 ^B	1.92 A	87.18 A	84.38 B

Table (5). The effect of amino acids on fruiting and fruit characteristics of olive trees cv.Manzanillo on two studied seasons (2021 and 2022)

Values sharing the same letter are not significantly different at the 5% significance level.

Fruiting and fruit character

Yield kg/tree

The data in Table (6) show that during the first experimental season, a glutamic spray at 50 mg/L significantly increased the yield to 18.3 kg/tree. In the 2^{nd} season, a mix of amino acid treatments led to a yield increase of 33.5 kg/tree. However, there were no significant differences between the control, proline, and GAPA treatments, all of which resulted in the lowest yields per tree in the 1^{st} season. In contrast, during the 2^{nd} season, the control treatment produced the lowest yield at 13.87 kg/tree.

This suggests that glutamic acid plays a key role in improving nutrient absorption and overall plant metabolism, leading to enhanced growth and productivity (**Shehata** *et al.*, **2021**). Glutamic acid is known to act as a precursor for other amino acids and has been shown to improve the efficiency of nitrogen use in plants, which likely contributed to the yield increase observed in this season (**Khalil & Hussein, 2020**).

Fruit weight (10 fruits)

From the results shown in Table 6, it is evident that the spray mix of amino acids had a significant effect. Notably, the 50 mg/L level produced the highest fruit weights, recording 47.07 g/10 fruits and 46.37 g/10 fruits in the 1st and 2nd seasons, respectively. In contrast, the lowest fruit weights were observed with the GAPA amino acid treatment, which resulted in 37.98 g/10 fruits and 41.17 g/10 fruits in the 1st and 2nd seasons, respectively.

These results demonstrate the positive role of amino acids in promoting fruit development, likely due to their involvement in various physiological processes, such as nutrient absorption, protein synthesis, and enzyme activation (Shehata *et al.*, 2021). Amino acids are known to improve overall plant vigor, which can directly contribute to increased fruit size and quality.

Oil %

The same trend in Table 6 also shows that the application of the amino acid mix at 50 mg/L had a significant positive effect, outperforming the control treatment. This treatment resulted in the highest oil content, with 23.7% in the 1^{st} season and 25.8% in the 2^{nd} season. On the other hand, the control treatment produced the lowest oil content in the fruits, with 21.60% in the 1^{st} season and 22.33% in the 2^{nd} season.

These findings align with previous research indicating the critical role of amino acids in plant growth, fruit development, and oil biosynthesis. Amino acids, as the building blocks of proteins, play an essential role in plant metabolism. They participate in the synthesis of enzymes and hormones that regulate various physiological processes, including nutrient uptake, stress tolerance, and secondary metabolite production (Shehata *et al.*, 2021). Specifically, amino acids such as proline, glutamic acid, and others can enhance the plant's capacity to assimilate nutrients and produce metabolites, which are crucial for oil synthesis in olive trees (Khalil & Hussein 2020). Amino acids promote the accumulation of oil in fruits by boosting the activity of enzymes involved in lipid biosynthesis. Additionally, these amino acids are known to enhance chlorophyll content, photosynthetic efficiency, and carbon fixation, all of which contribute to improved fruit quality and increased oil content (Abdel-Mawgoud *et al.*, 2020).

Treatment	Yi	eld	Fruit (10 f	weight Truit)	Oil %		
	1 st	2 nd	1 st	2 nd	1 st	2 nd	
Control	6.22 ^C	13.87 ^C	40.37 ^{BC}	42.80 ^{BC}	21.60 ^d	22.33 ^D	
Glutamic	18.30 ^A	23.17 ^B	41.26 ^B	44.73 ^{AB}	23.47 ^{AB}	24.67 ^B	
Proline	7.50 ^C	15.50 ^C	39.87 ^{BC}	42.73 ^{BC}	22.86 ^{BC}	23.40 ^c	
Gapa	7.63 ^C	22.33 ^B	37.98 ^C	41.17 ^C	22.40 ^{CD}	22.83 ^{CD}	
Mix	12.18 ^B	33.50 ^A	47.07 ^A	46.37 ^A	23.70 ^A	25.80 ^A	

Table (6). The effect of amino acids on yield (Kg/ tree), fruit weight (g/10 fruits) and oil % of olive trees cv. Manzanillo on two studied seasons (2021 and 2022)

Values sharing the same letter are not significantly different at the 5% significance level.

Chemical contents in olive leaves

Total amino acid content (g/100 g)

The results for amino acid content presented in Table (7) show significant variation among the different treatments. The application of the amino acid mixture resulted in a notable increase in total amino acid content during both study seasons, ranging from 0.0625 to 0.0580. The control treatment, however, exhibited the lowest amino acid content values, while the other treatments yielded intermediate values in both seasons.

Total carbohydrate (%)

Data in Table (7) showed the effect of foliar application of different types of amino acids on carbohydrate (%) content in olive leaves. the control treatment recorded statistically the highest carbohydrate content (33.00- 34.50 %) in the two investigated seasons. Meanwhile, the lowest carbohydrate content was achieved by the mixture of amino acid treatment (17.22- 15.71 %) in the 1st and 2nd seasons, respectively.

The accumulation of carbohydrate seems to be one of the survival mechanism for plants grown under salt stress (**El Yamani and Cordovilla, 2024**). In abiotic reactions, carbohydrates serve as substrates for osmotic regulation, protection, and energy generation that connect growth development and carbon status (**Diniz** *et al.*, **2020**). In addition, **Tattini** *et al.*, (2006) showed that, carbohydrates increased as the external NaCl concentration was increased.

Treatment	Total amino	acid g/100 g	Total carbohydrate %		
reatment	1 st	2 nd	1 st	2 nd	
Control	0.0400 d	0.0435 c	33.00 a	34.50 a	
Glutamic	0.0485 b	0.0500 b	19.82 d	19.92 c	
Proline	0.0405 d	0.0455c	28.54 b	25.92 b	
Gapa	0.0425 c	0.0460 c	22.02 c	20.22 c	
Mix	0.0625 a	0.0580 a	17.22 e	15.71 d	

Table (7) the affect of amino acids on total o	carbohydrate and amino acid content in olive leaves.
Table (7) the chect of annua actus on total c	car bonyur ate and annu actu content in onve reaves.

Values sharing the same letter are not significantly different at the 5% significance level.

4. Conclusion

Based on obtained data in this research, it could be noted that the foliar application of Glutamic amino acid alone significantly was the best treatment compared to other free amino acid during this study and mixture of free amino acid enhance the vegetative growth and productivity of olive trees (Manzanillo cv.).

References

A.O.A.C., (1998). Association of official Agricultural Chemists. Official methods of Analysis, 14 ed., P.O. Box uso, Benjamin franklin station, Washington, D.C., pp: 832.th

Abdel-Mawgoud, A. M. R., Tantawy, A. S., Hafez, M. M. and Habib, H. A. M. (2020). The role of amino acids in improvement in plant growth, yield, and some physiological aspects of snap beans grown under high temperature conditions. Plant Physiology and Biochemistry, 82, 303-311.

Aberg, B. (1961). Nucleic acids and proteins in plants. Encycl. Plant Physiol., (14) Spriger Verlag, Berlin.

Ahmad, P. and Sharma, S. (2021). Role of proline and glutathione in combating oxidative stress under salt stress conditions. Journal of Plant Growth Regulation, 40(1), 159-170.

Ali, Q., Shabbir, G. and Khan, M. I. R. (2021). "Exogenous application of amino acids and their role in mitigating salt stress in plants: A review." Agronomy, 11(2), 309.

Al-Maaitah, M. I., Shatnawi, M. A. and Al-Mustafa, W. A. (2017). "Effects of salinity stress on growth and nutrient uptake of olive (*Olea europaea* L.) cultivars." Journal of Agricultural Science and Technology, 19(3), 701-710.

Almutairi, K. F.; Saleh, A. A.; Ali, M. M.; Sas-Paszt, L.; Abada, H. S. and Mosa, W. F. A. (2022). Growth Performance of Guava Trees after the Exogenous Application of Amino Acids Glutamic Acid, Arginine, and Glycine. Horticulturae, 8, 1110.

Ben Ahmed, C., Ben Rouina, B. and Boukhris, M. (2020). "Foliar application of amino acids affects hormonal regulation and stress tolerance in olive trees under saline conditions." Journal of Plant Nutrition and Soil Science, 183(4), 501-512.

Brosnan, J. and Brosnan, M. E. (2013). Glutamate: A truly functional amino acid. Amino Acids, 45, 413–418.

Calvo, P.; Nelson, L. and Kloepper, J. (2014). Agricultural uses of plant biostimulants. Plant Soil, 383, 3–41.

Davies, D.D. (1982). Physiological aspects of protein turn over. Encycl. Plant Physiol., 45: 481-487.

Di Vaio, C., Petriccione, M. and Amoroso, G. (2024). "Recent Advances in Amino Acid-Based Biostimulants for Olive Cultivation in Saline Environments." Journal of Plant Nutrition and Soil Science, 187(1), 34-46.

Diniz A L, da Silva D I R, Lembke C G, Costa M D-B L, ten-Caten F, Li F, Vilela R D, Menossi M, Ware D, Endres L. and Souza G. M. (2020). Amino Acid and Carbohydrate Metabolism Are Coordinated to Maintain Energetic Balance during Drought in Sugarcane. International Journal of Molecular Sciences. 21(23):9124.

Dubois, M., Smith, F., Gilles, K., Hammilton, J. K. and Robers, P. A. (1956). Colorimetric method to determination of sugars and related substances. Anel. Chem. ,28 (3): 350-356.

Duncan, D.B. (1955). Multiple range and multiple, F-test Biometrics, 11: 1-42.

Eisaa R.A., Merwadb M.A., Mostafab E.A.M., Salehb M.M.S. and Ashourb N.E. (2023). The Impact of Spraying Selenium, Glutamic Acid and Seaweed Extract on Growth, Productivity, Physical and Chemical Fruit Properties of Banana. Egypt. J. Chem. Vol. 66, No. 1 pp. 121 – 128.

El-Badawy, H. E. M. (2019). Effect of Spraying Amino Acids and Micronutrients as Well as their Combination on Growth, Yield, Fruit Quality and Mineral Content of Canino Apricot Trees. J. Plant Production, Mansoura Univ., 10 (2): 125 - 132.

El- Bolok, T. Kh. and Kasem, M.S.M. (2023). Effect of Foliar Application with Chitosan and Amino Acids on Growth, Flowering, Yield and Fruit Quality of Aggizi Olive Trees Under Qena Governorate Conditions. Horticulture Research Journal, 1(1), 52:67.

Ekşi H.S. and Sönmez I. (2022). Effects of amino acid applications on yield, growth and mineral nutrition of greenhouse tomato. J. Elem., 27(3): 545-557.

El-Esawi, M. A., Al-Ghamdi, A. A. and Ali, H. M. (2020). "Exogenous application of cysteine and methionine improves salt stress tolerance in soybean by modulating antioxidant defense and stress-responsive genes." Environmental and Experimental Botany, 176, 104087.

El-Esawi, M. A. and Ali, H. M. (2022). "Influence of amino acids on secondary metabolites and stress tolerance in plants." Journal of Plant Interactions, 17(1), 102-113.

El-Tayeb, M. A. (2021). "Amino acids and their role in the biosynthesis of compatible solutes and polyamines in stress tolerance." Plant Stress, 5, 100046.

El Yamani M. and Cordovilla M. P. (2024). Tolerance Mechanisms of Olive Tree (Olea europaea) under Saline Conditions. Plants, 13(15):2094.

Fariduddin, Q., Ali, A., and Hayat, S. (2021). "Amino acids as modulators of plant hormone signaling pathways under stress conditions." Journal of Plant Physiology, 263, 153489.

Farooq, M., Hussain, M. and Wakeel, A. (2020). "Glycine betaine and proline enhance tolerance to salt stress in wheat." Plant Physiology and Biochemistry, 150, 452-462.

Feng, H., Fan, X., Miller, A. J. and Xu, G. (2020). Plant nitrogen uptake and assimilation: regulation of cellular pH homeostasis. Journal of experimental botany, 71(15), 4380–4392.

Ghars, M. A., Hosni, K. and Guizani, M. (2022). "Role of Exogenous Amino Acids in Enhancing Salt Tolerance and Olive Oil Quality." Plant Physiology and Biochemistry, 182, 253-262.

Goss, J.A. (1973). Amino acid synthesis and metabolism. In Physiology of. Plants and their cells. Pergamon Press, Inc., New York; p.202.

Hamed, S., Ali, M. and El-Rawy, M. (2021). "Effects of amino acid treatments on ion homeostasis and sodium toxicity in olive trees under saline stress." Journal of Horticultural Science and Biotechnology, 96(3), 306-315.

Hasanuzzaman, M., Bhuyan, M. H. M. B., Raza, A. and Fujita, M. (2020). "Reactive oxygen species and antioxidant defense in plants under abiotic stress: Revisiting the crucial role of a universal defense regulator." Antioxidants, 9(8), 681.

Hassan, F. A. S. and Ali, E. F. (2022). "Amino acid-mediated modulation of salt stress tolerance in olive trees and its effect on oil quality." Plants, 11(1), 67.

Hu, B., Jin, J., Guo, A. Y., Zhang, H., Luo, J. and Gao, G. (2015). GSDS 2.0: an upgraded gene feature visualization server. Bioinformatics, 31(8), 1296-1297.

Iqbal, N., Umar, S. and Khan, N. A. (2021). "Osmolytes and plant adaptation to salinity: Focus on the role of proline, glycine betaine, and arginine." Environmental and Experimental Botany, 188, 104485.

Khalil, H. A. and Hussein, M. A. (2020). The effect of foliar application of amino acids and micronutrients on olive productivity and fruit quality. Agronomy, 10(3), 123-130.

Khan, M. I. R. and Müller, M. (2022). "Modulation of plant hormone signaling pathways by amino acids in response to abiotic stresses." International Journal of Molecular Sciences, 23(4), 1786.

Kumar, A. and Singh, A. (2020). "Role of amino acids in the synthesis of stress-responsive metabolites and osmoprotectants in plants." Frontiers in Plant Science, 11, 1055.

Liu, S., Li, Y. and Zhang, C. (2022). "Amino acids and secondary metabolite production in plants under stress conditions: Insights into regulatory mechanisms." Metabolites, 12(4), 357.

Lodolini, E. M., Gucci, R. and Servili, M. (2019). "Olive tree and olive oil production: Strategies to improve sustainability and quality." Journal of Horticultural Science & Biotechnology, 94(5), 485-497.

Mervat, S. H. S., Abdelhamid, M. T. and Schmidhalter, Urs (2015). Effect of foliar application of aminoacids on plant yield and some physiological parameters in bean plants irrigated with seawater. Acta Biológica Colombiana, 20 (1), 141-152.

Mosleh, Z. and Soltani, F. (2023). "Improving Olive Tree Productivity under Salinity Stress: Foliar Application of Amino Acids Combined with Micronutrients." Journal of Environmental Biology, 44(4), 719-727.

Nafea A. M. A. and Al-Janabi A. M. I. (2021). Effect of Foliar Application with Kinetin and Amino Acids in the Vegetative Growth and Chemical Content of Young Olive Trees cv. & quot; K18". Annals of the Romanian Society for Cell Biology, 10067–10076.

Nogueira, F. D., Martins, M. L., & Gomes, M. (2022). "Salt stress effects on physiological traits and productivity of olive trees." Plant Stress, 5, 100063.

Orlandi, F., Bonofiglio, T. and Romano, B. (2021). "Proline and glycine betaine accumulation in olive (*Olea europaea* L.) trees exposed to salt stress and the impact on oil yield." Acta Physiologiae Plantarum, 43, 1-12.

Psarras, G., Manolikaki, I., Dareioti, M., Digalaki, N., Sergentani, C., Barbopoulou, E. and Koubouris, G. (2024). Effect of amino acids application on flowering, vegetation, yield, and oil of olive (*Olea europaea* L.) variety 'Koroneiki.' Journal of Plant Nutrition, 47(13), 2057–2069.

Senedecor, C.W. and Cochran, W.G. (1990). Statistical methods 7th ed The low State Univ. Press. Ames Iowa USA. pp: 593.

Shehata, S. A., Taha, H. A. and Gad, N. (2021). Application of amino acids in agriculture: An overview. Journal of Plant Nutrition, 44(3), 435-445.

Tattini, M., Gucci, R., Romani, A., Baldi, A., Everard, J. (2006). Changes in non-structural carbohydrates in olive (Olea europaea) leaves during root zone salinity stress. Physiologia Plantarum. 98. 117 – 124.

Thon M., Maretzki A., Korner E. and Soki W.S. (1981). Nutrient uptake and accumulation by sugar cane cell culture in relation to growth cycle. Plant Cell Tiss Org.; (1):3-14.

Wang, X., Chen, X. and Xu, Y. (2022). "Glutamine application enhances ion homeostasis and alleviates sodium toxicity in salt-stressed olive trees." Journal of Plant Physiology, 260, 153435.

Wilde, S.A.; Corey, R.B.; Layer, J.G. and Voigt, G.K. (1985). Soils and Plant Analysis for tree culture. Oxford, Publishing Co., New Delhi, pp. 96-106.

Xu, Z., Ma, J., Lei, P., Qian W. and Xiaohai F. (2020). Poly-γ-glutamic acid induces system tolerance to drought stress by promoting abscisic acid accumulation in Brassica napus L. Sci. Rep., 10, 252.

Yemm, E. W. and Cocking, E. C. (1995). The determination of amino acids with ninhydrin. Analyst. 80: 208-214.

Zayed, O., Hewedy, O. A., Abdelmoteleb, A., Ali, M., Youssef, M. S., Roumia, A. F., Seymour, D. and Yuan, Z. C. (2023). Nitrogen Journey in Plants: From Uptake to Metabolism, Stress Response, and Microbe Interaction. Biomolecules, 13 (10), 1443.

Zhang, H. and Jiang, M. (2021). "The role of amino acids in the regulation of plant hormones and stress responses." Journal of Plant Growth Regulation, 40(1), 190-202.

Zhang, J. and Zhou, H. (2020). "The role of cysteine and asparagine in maintaining ion homeostasis and stress tolerance in plants." Plant Cell Reports, 39(6), 759-770.

Zrig, A., Hannachi, H. and Ben Salem, F. (2023). "Impact of foliar application of amino acids on growth and yield of olive trees under salt stress conditions." Agronomy Journal, 115(7), 1021-1033.



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